

# ENHANCEMENT OF OSTEOGENIC CELL RESPONSE ON BIOMIMETIC IMPLANT COATINGS

M. Gorbahn<sup>1,2</sup>, M. Klein<sup>3</sup>, M. Lehnert<sup>1</sup>, D. Brüllmann<sup>4</sup>, I. Köper<sup>2</sup>, B. Al-Nawas<sup>3</sup>, M. Veith<sup>1</sup>

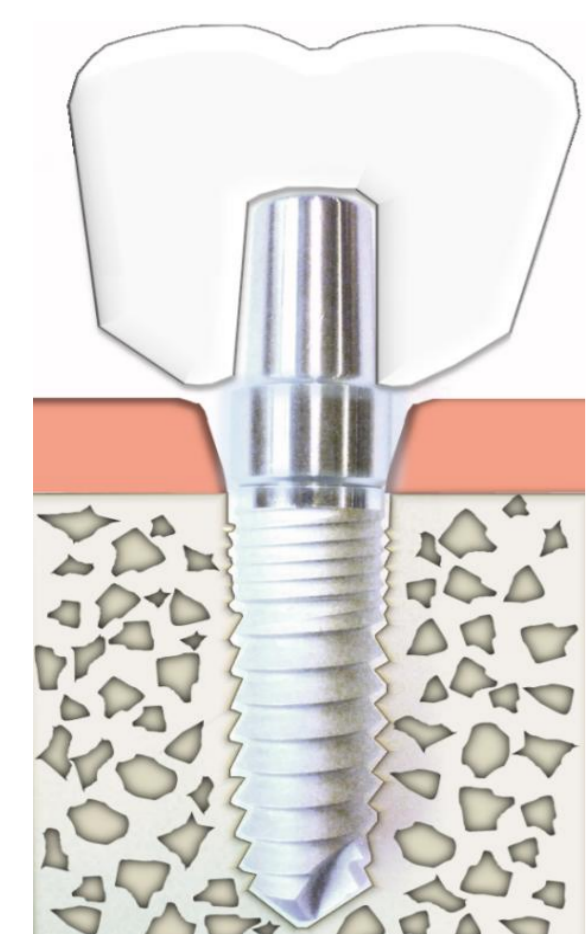
<sup>1</sup> University of Applied Science Gelsenkirchen, Department of Applied Natural Science, 45665 Recklinghausen, Germany;

<sup>2</sup> Max Planck Institute for Polymer Research, Department of Materials Science, 55128 Mainz, Germany;

<sup>3</sup> University Medical Center of the Johannes Gutenberg University Mainz, Center of Oral and Maxillofacial Surgery, 55101 Mainz, Germany;

<sup>4</sup> University Medical Center of the Johannes Gutenberg University Mainz, Center of Oral Surgery, 55101 Mainz, Germany

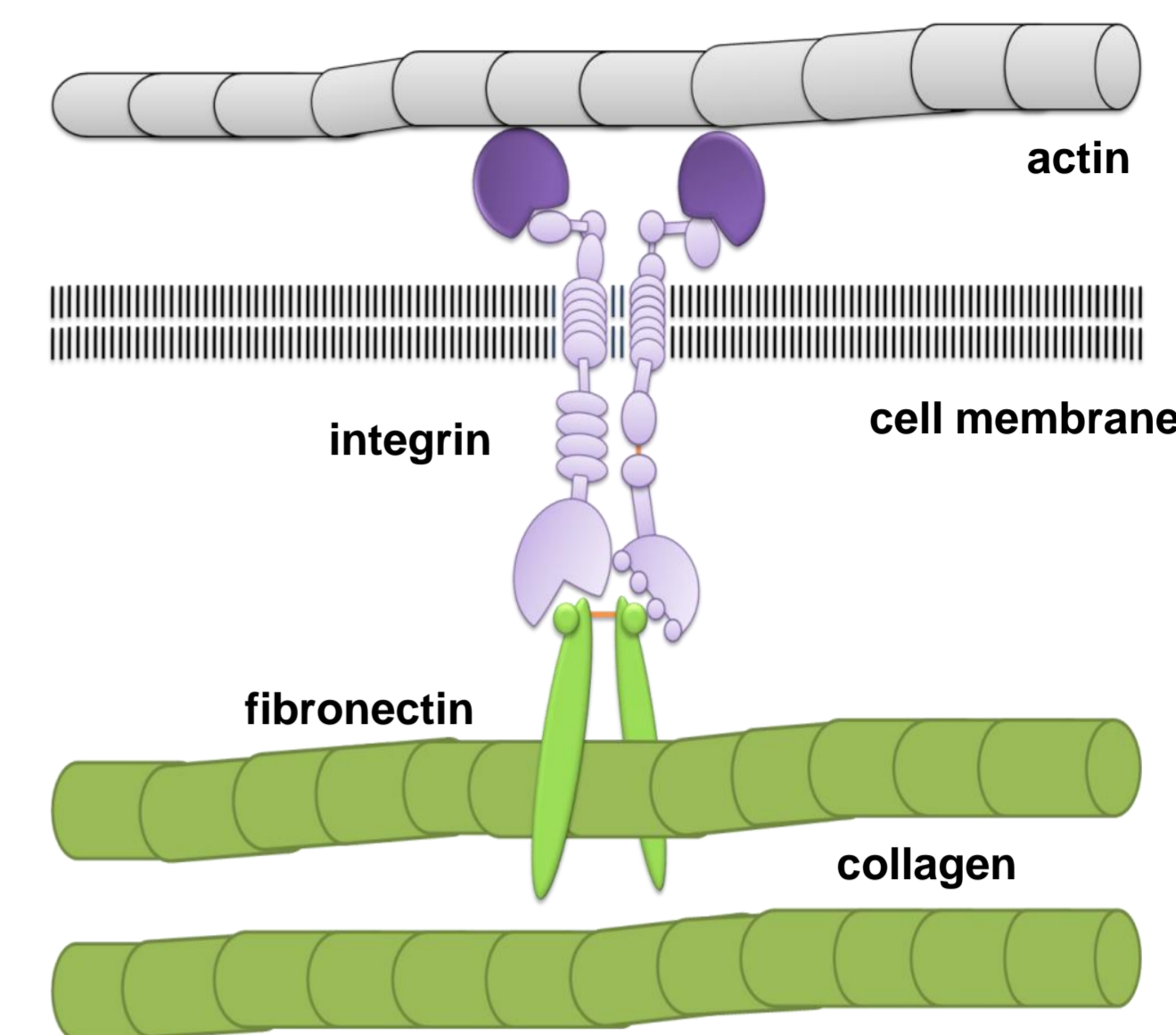
## Introduction



Titanium has been used over the last decades as material for bone-anchored implants. However, there are still cases of implant failure. A possible improvement can be the coating of the implant surface with biological coatings using proteins or peptides.

Fibronectin (FN) is an adhesion molecule from the extracellular matrix (ECM). In contact with cellular integrin receptors, FN forms stable connections between the ECM and the cells. Therefore, FN is an interesting material for biomimetic implant coatings.

Biotinylated FN (bFN) has been immobilized on TiO<sub>2</sub> surfaces in a stable and flexible way with preservation of biological activity using a Biotin-Streptavidin system.

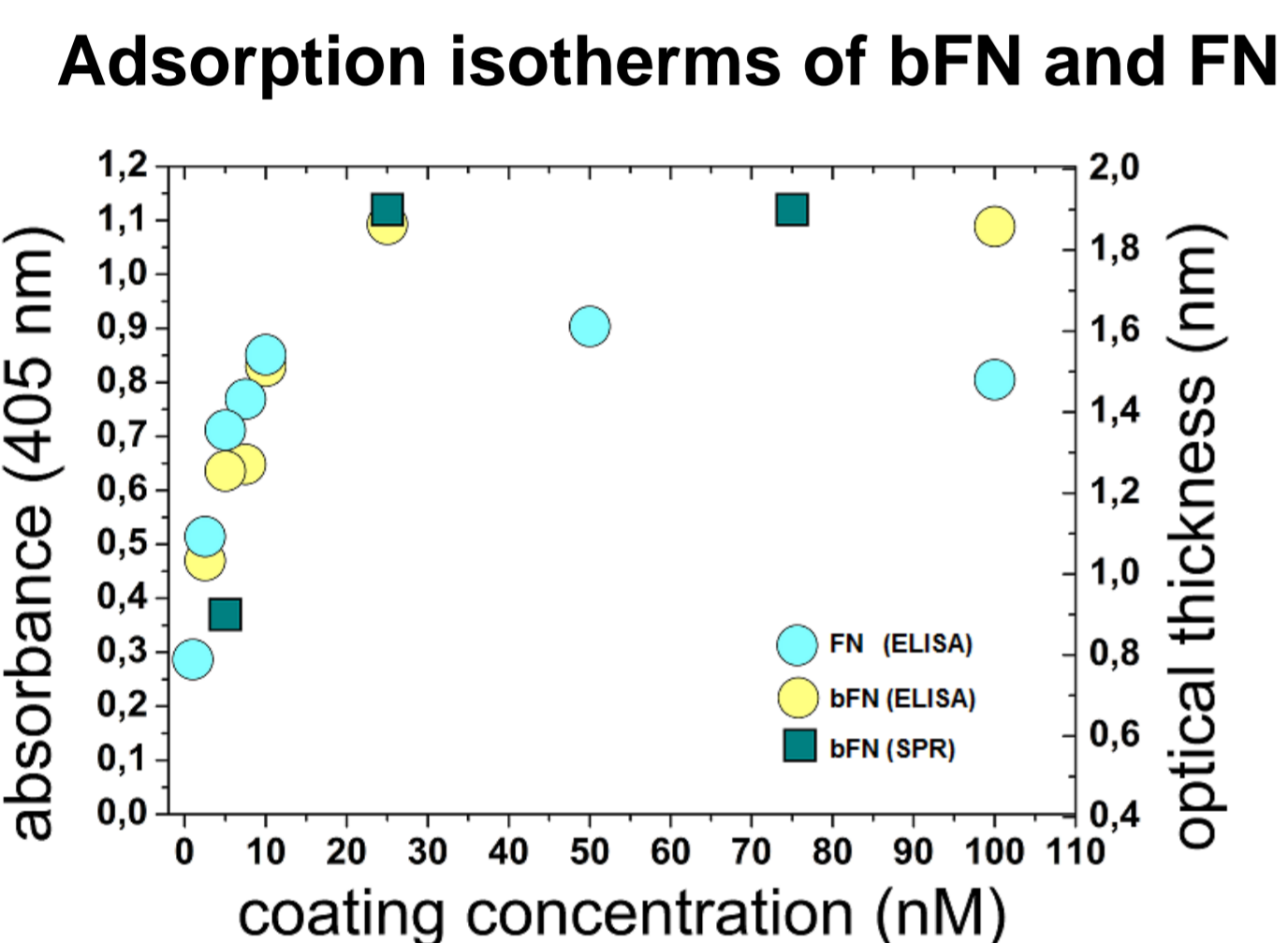


## Methods

1. Biomimetic coating of bFN over a Biotin-Streptavidin system *via self assembly*
2. Investigation of adsorption conditions by **SPR** (surface plasmon resonance) spectroscopy
3. Investigation of physicochemical properties by **ELISA** (enzyme linked immunosorbent assay)
4. Investigation of cell proliferation and differentiation by **RT-qPCR** (Quantitative Real Time-polymerase chain reaction)
5. **Cell adhesion** tests with HOBs (human osteoblasts) cells

## Results

### Adsorption behaviour & surface density

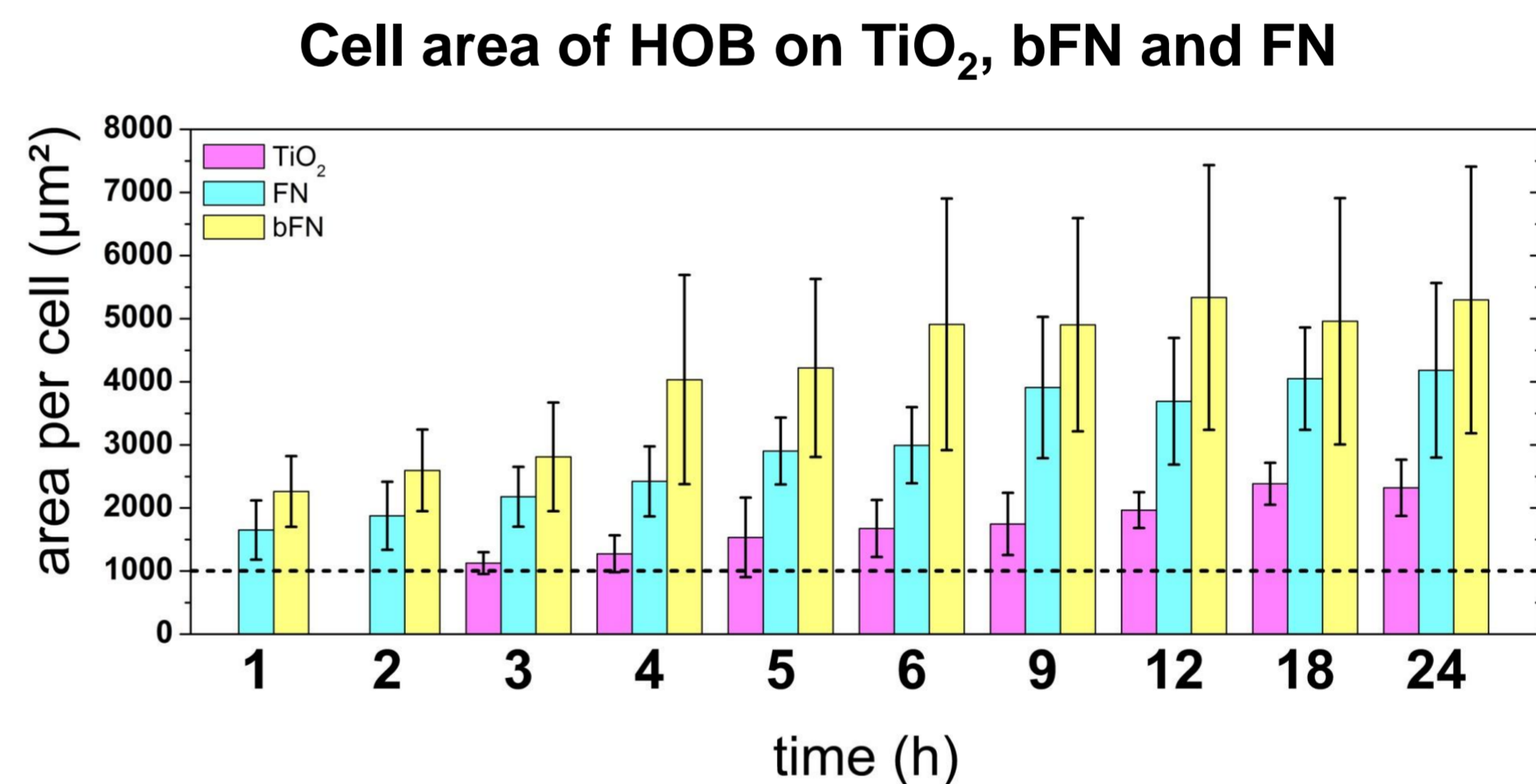


### Saturation limit and Surface density

	Saturation Limit	Surface Density
FN	10 nM	0.36 µg/cm <sup>2</sup>
bFN	25 nM	0.88 µg/cm <sup>2</sup>

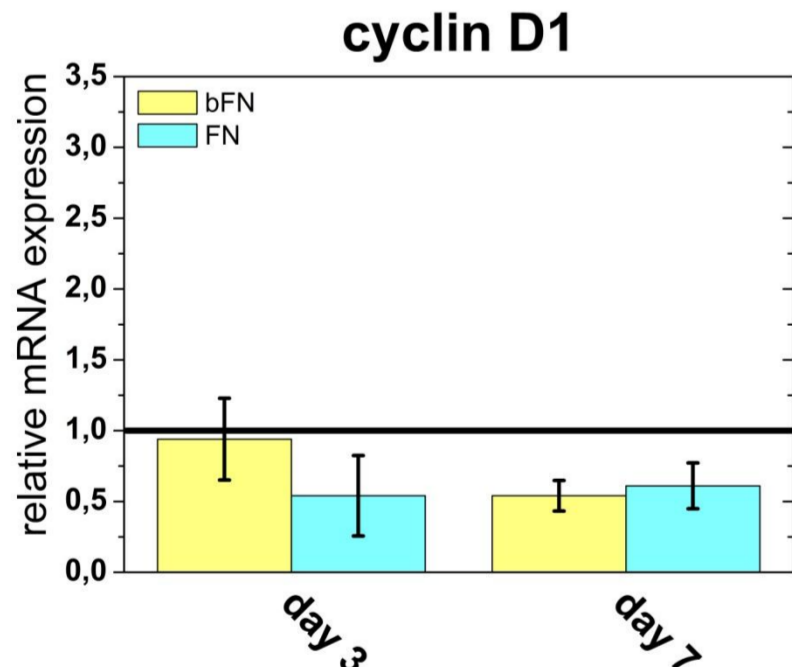
→ Saturation limits were detected at coating concentrations of 25 nM for bFN and 10 nM for FN with surface densities at 0.88µg/cm<sup>2</sup> for bFN and at 0.36µg/cm<sup>2</sup> for FN.

### Cell Adhesion



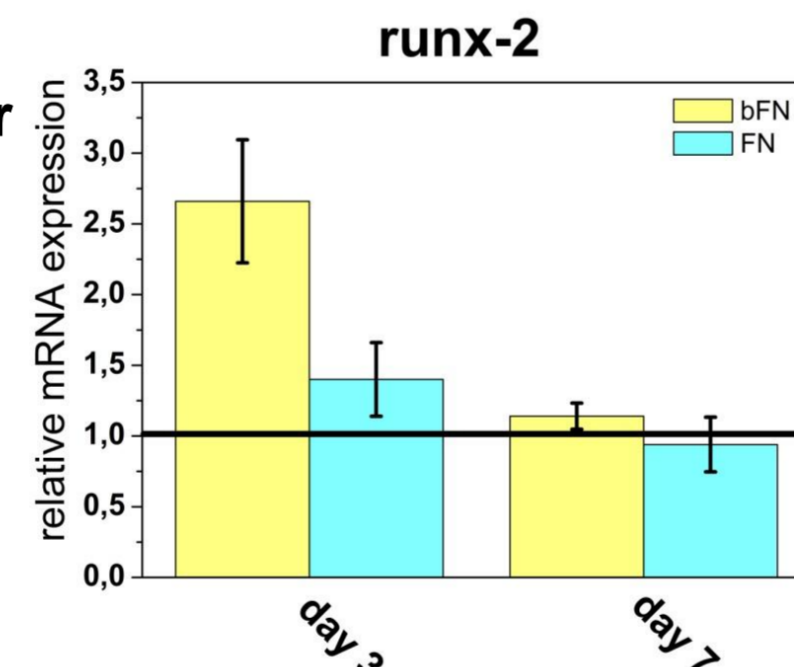
### Cell Proliferation

#### Cell proliferation and differentiation of HOB on bFN and FN (normalized to 18S and compared with TiO<sub>2</sub>)



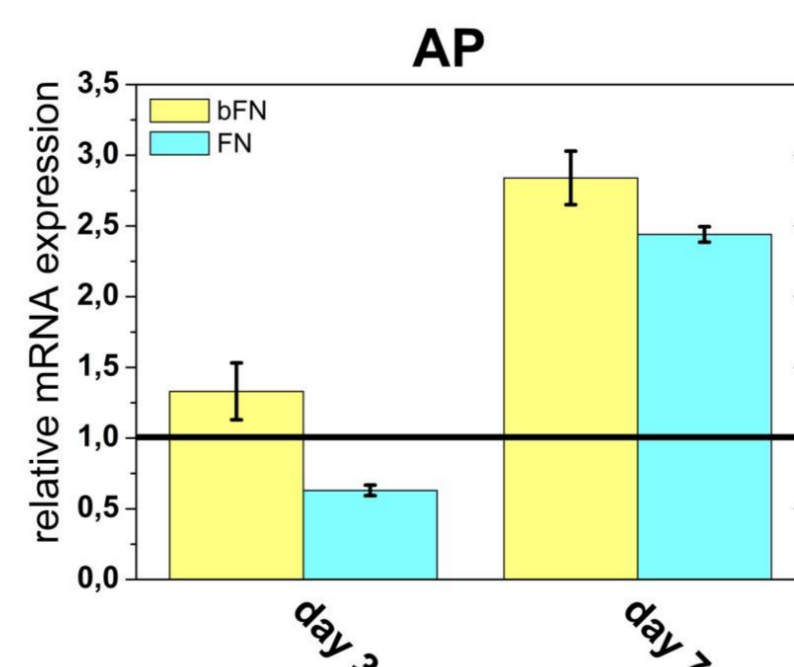
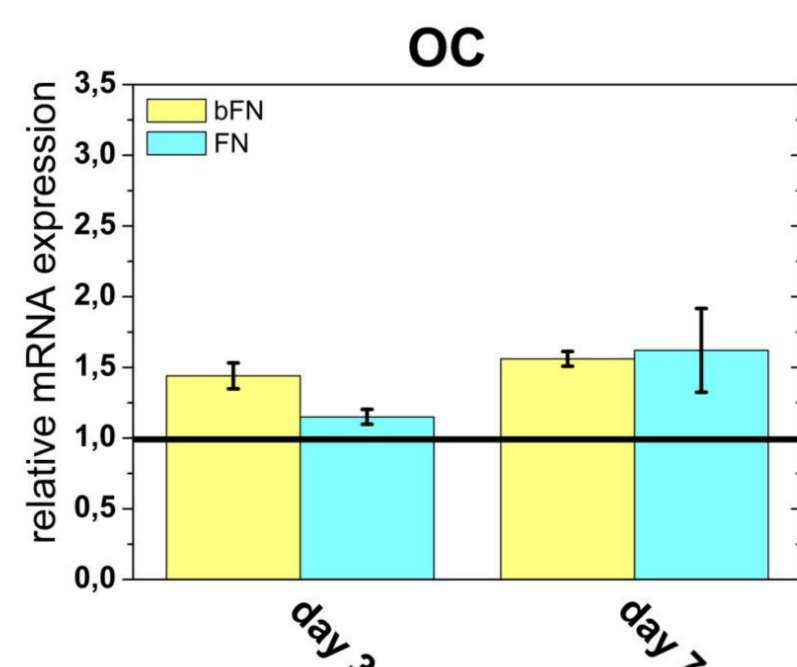
**Proliferation marker**  
Cyclin D1

**Transcription factor**  
Runx-2



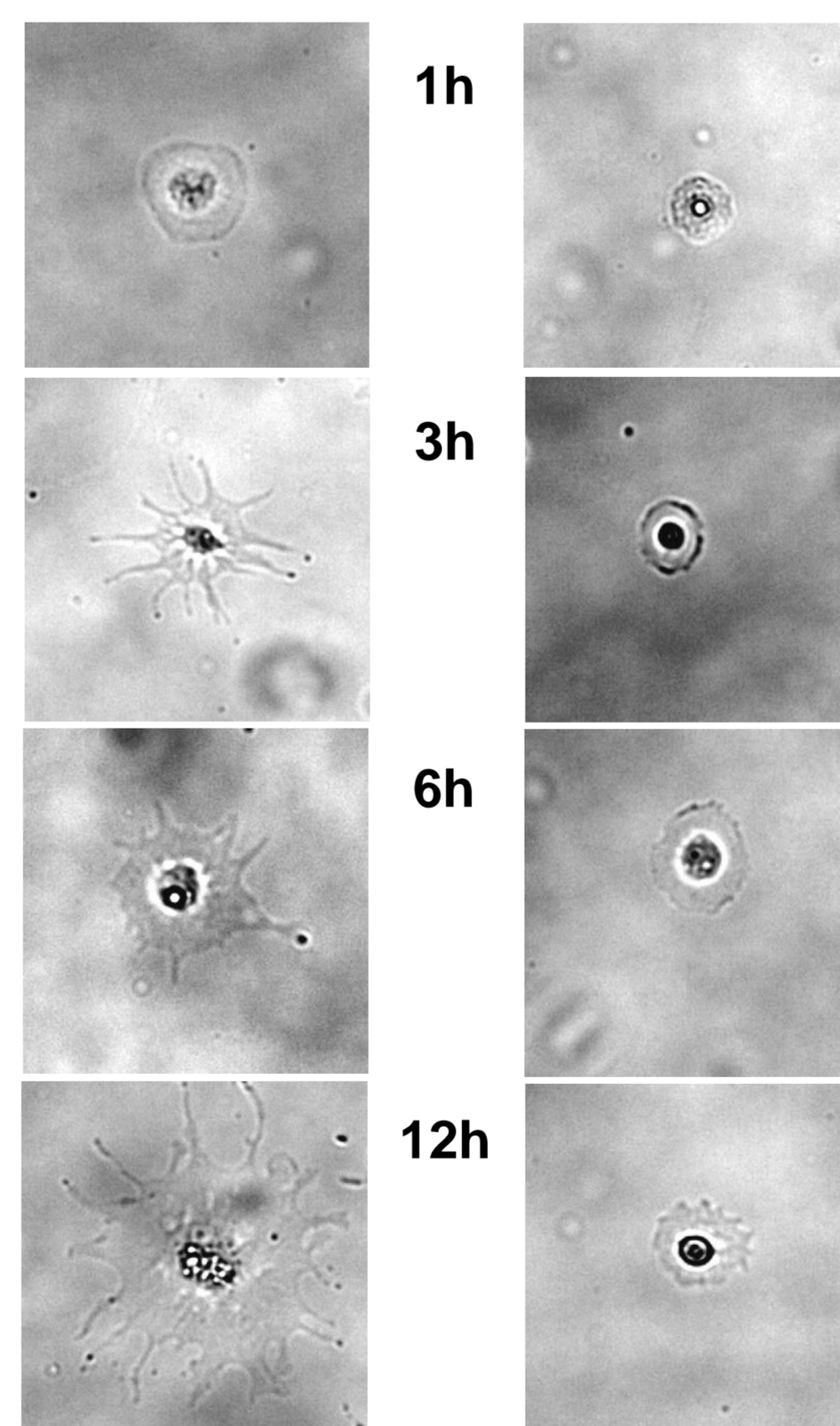
**Osteoblast-specific marker**  
AP (Alkaline Phosphatase)  
OC (Osteocalcin)

- Decreased cyclin D1 expression after 7 days.
- Early response of runx-2 expression after 3 days.
- Enhancement of OC and AP expression after 7 days.

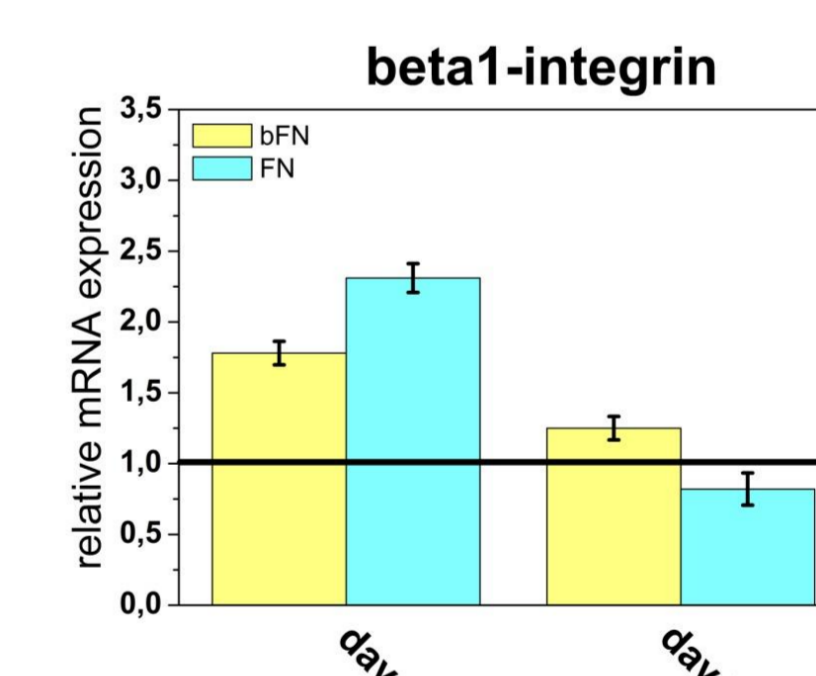


### Cell Differentiation

#### Initial cell morphology of HOB on bFN (left) and TiO<sub>2</sub> (right)



#### Cell adhesion of HOB on bFN and FN (normalized to 18S and compared with TiO<sub>2</sub>)



- Cell adhesion as a function of cell area was enhanced on bFN and FN coatings.
- Initial cell morphology showed faster cell growth and greater cell spreading on bFN compared to TiO<sub>2</sub>.
- Expression of β1-integrin was increased on bFN and FN after 3 days and normalized after 7 days.

## Conclusion

- Surface density and saturation limit were two-fold higher for specific immobilized bFN than for unspecific adsorbed FN [1].
- Adsorption isotherms observed with ELISA confirmed previous SPR measurements [2].
- Biomimetic coatings with bFN and FN enhanced osteogenic cell response and triggered early cell differentiation after 3 days.
- Due to this, cell proliferation decreased after 7 days [3].
- Cell adhesion tests indicated a promotion of cell area and β1-integrin expression on bFN and FN.
- Initial cell morphology showed also an enhancement of cell spreading on bFN compared to TiO<sub>2</sub> [3].

## Acknowledgement

The authors thank Dr. Beyer, Prof. Dr. Loidl-Stahlhofen, and S. Tröder for helpful comments and discussion. Special thanks go to Dr. U. Ritz, A. Ackermann, L. Groothusen, and U. Zerfaß, for excellent technical assistance. This project is supported by a grant from the FH<sup>3</sup> BMBF program through the cooperative project "Bionanofunktionalisierte Oberflächen" (FKZ1775X05).

## References

- [1] Gorbahn M, Lehnert M, Beyer A, Köper I, Veith M; submitted to Biomedical Materials 2009
- [2] Lehnert M, Gorbahn M, Rosin C, Köper I, Knoll W, Veith M; in preparation
- [3] Gorbahn M, Klein M, Lehnert M, Brüllmann M, Wagner W, Al-Nawas B, Veith M; submitted to Clinical Oral Investigations 2009

## Contact: Miriam Gorbahn



**Research group: Prof. Dr. Michael Veith**  
Tel.: 02361/915-484 (Fax -486)  
E-mail: michael.veith@fh-gelsenkirchen.de  
University of Applied Science, Gelsenkirchen  
Department of Applied Natural Science  
45665 Recklinghausen